

V79 COLONY FORMING ASSAY

Experiment Name : $^{125}\text{IUdR}$ + 50-200 ug/ml MEA; **Exp. # :** 1; **Investigator:** A.Bishayee
Date: 05/14/98

1. Set the rocker-roller at 37°C incubator, set the Coulter Counter, wash cells (from 75 cm² flask, subcultured 1:2, 24h before) with PBS, trypsinize cells, resuspend in 7 ml MEMB, pass five times through 3 cc syringe with 21 gauge needle, perform cell count by transferring 100 ul in Coulter cup containing 20 ml isotone (Coulter balanced electrolyte solution)
2. Dilute to ~4,00,000 cells/ml in MEMB (final volume 11 ml) [Actual count : 415600 cells/ml]
3. Transfer 1 ml of cell suspension into ten 12 ml tubes (Falcon plastic test tube, 17x100 mm) labeled 1-10 both on cap and wall
4. Roll the tubes for 3-4 h at 37°C, 5% CO₂ Date/Time: 05/14/98; 4-15 p.m.
5. Prepare MEMB containing radioactivity in hood
1.95 µl $^{125}\text{IUdR}$ (prepared on 04/09/98) + 2.5 ml MEMB
6. After 3-4 h, remove test tubes from roller and add MEMB with or without radioactivity according to Table below. Date/Time: 05/14/98; 7-45 p.m.

Tube #	$^{125}\text{IUdR}$ uCi/ml	Cells in MEMB (ml)	MEMB (ml)	MEMB+ $^{125}\text{IUdR}$ (ml) [1.0 uCi/ml]	MEMA+ MEA (200 ug/ ml) (ml)	MEMA (ml)	MEA Conc. (ug/ml)
1	0	1.0	1.0	0	0	2.0	0
2	0	1.0	1.0	0	0.5	1.5	50
3	0	1.0	1.0	0	1	1	100
4	0	1.0	1.0	0	1.5	0.5	150
5	0	1.0	1.0	0	2.0	0	200
6	0.02	1.0	0.96	0.04	0	2.0	0
7	0.02	1.0	0.96	0.04	0.5	1.5	50
8	0.02	1.0	0.96	0.04	1	1	100
9	0.02	1.0	0.96	0.04	1.5	0.5	150
10	0.02	1.0	0.96	0.04	2.0	0	200

7. Return test tubes to roller for 12 h, increase the elevation angle of the roller.

Date/Time: 05/14/98; 8-00 p.m.

8. While test tubes are rolling label 40 (4x10) gamma-tubes (13 X 100 mm VWR glass test tube)
9. After ~12 h incubation period, remove tubes and centrifuge at 2000 rpm at 4°C for 10 min (precooled centrifuge). Date/Time: 05/15/98; 9-60 a.m.
10. During centrifugation, move roller to 10°C and obtain ice MEMA = 10.89 ml
11. Prepare 11 ml of 200 ug/ml MEA in MEMA, put on ice MEA (200ug/ml) = 0.11 ml
12. Remove buckets from centrifuge and carefully remove 100 µl of supernatant and place in prelabeled gamma-tube.
13. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
14. Centrifuge tubes for 10 min at 2000 rpm, 4°C
15. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
16. Centrifuge tubes for 10 min at 2000 rpm, 4°C
17. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
18. Centrifuge tubes for 10 min at 2000 rpm, 4°C
19. Decant supernatant, click tubes, vortex, resuspend in 2 ml ice cold MEMA containing 0 and/or 200 ug/ml MEA as per Table. Keep on ice!
20. Transfer tubes to roller at 10°C for 72 h. Date/Time: 05/15/98; 11-00 a.m.
21. Transfer 10 ul supernatant in three sets of tubes containing small pieces of tissue paper from 100 ul supernatant removed earlier and count them for radioactivity Date/Time: 05/26/98; 11-30 a.m.
21. After 72 h, remove tubes and place on ice, add 8 ml ice cold wash MEMA. Date/Time: 05/18/98; 1-00 p.m.
22. Centrifuge tubes for 10 min at 2000 rpm, 4°C (precooled centrifuge)
23. Labeling and preparation of dilution tubes and colony dishes
 - load 57 60 mm petri dishes with 4 ml MEMA
 - load 30 T-tubes with 4.5 ml MEMA and label them 1.2, 1.3, 1.4, 2.2, 2.3, 2.4, X.2, X.3, X.4, etc.
24. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
25. Centrifuge tubes for 10 min at 2000 rpm, 4°C
26. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
27. Centrifuge tubes for 10 min at 2000 rpm, 4°C
28. Decant supernatant, click tubes, vortex, resuspend in 2 ml wash MEMA, pass five times through 3 cc syringe with 21 gauge needle
29. Determine cell concentration by transferring 100 µl to Coulter cup
30. Vortex tube, transfer 0.5 ml into dilution tube X.4, vortex tube X.4 and transfer 0.5 ml to tube X.3, vortex tube X.3 and transfer 0.5 ml to tube X.2 and vortex. Keep tubes on ice.
31. Transfer 1 ml from dilution tubes into dishes labeled X.2, X.3, X.4 (in triplicate). Only X.2 should be seeded for control T-tubes.

32. Transfer 300 μ l of cell suspension (in triplicate) to gamma tubes for each tube

33. Incubate petridishes for 1 week

34. Count gamma tubes for radioactivity

Date/Time : 05/26/98; 12-00 noon

35. After 1 week, wash colonies 3 times with normal (1X) saline, and 2 times with methanol.

Stain colonies with crystal violet

36. Count colonies (50 or more cells). There must be between 25 and 250 colonies for the flask to be a valid data point.

Expt #1

05/14/98

$$\begin{aligned} \text{Initial Cell Count} &= 6330, 6320, 6320 \\ \text{Avg. Cell Count} &= 6323.3 \\ \text{Cell Conc.} &= 6323.3 \times 400 \\ &= 2529333.3 \text{ Cells/ml} \end{aligned}$$

For dilution,

$$\begin{aligned} \text{Vol of Cell Suspension taken} &= \frac{4400000}{2529333.3} \\ &= 1.73 \text{ ml} \end{aligned}$$

Take 1.73 ml of cells + 9.27 ml MEMB = 11 ml.

After dilution,

$$\begin{aligned} \text{Final cell count} &= 1003, 1058, 1056 \\ \text{Avg. cell count} &= 1039 \\ \text{Cell conc.} &= 1039 \times 400 \\ &= 4,15,600 \text{ cells/ml} \end{aligned}$$

Expt # 1

05/14/98

Preparation of ^{125}I UDR in MEMB.

Prepare 2.5 ml of $1 \mu\text{Ci}/\text{ml} = 2.5 \mu\text{Ci}$ required

Stock

ON 04/09/98 = $1.93 \mu\text{Ci}/\text{ml}$

ON 05/14/98 = 1.93×0.667
= $1.28 \mu\text{Ci}/\text{ml}$.

Stock required = $\frac{2.5}{1.28} = 1.95 \text{ ml}$.

- ① Take 2.5 ml of MEMB
- ② Add 1.95 ml of ^{125}I UDR

cpm for 10 µl medium onto tube

1	1.00		34	266
2	1.00	40	47	283
3	1.00			
4	1.00	1M	25	261
5	1.00		47	274
6	1.00		30	288
7	1.00	2M	64	287
8	1.00		40	280
9	1.00		49	300
10	1.00	3M	56	326
11	1.00		48	284
12	1.00		33	289
13	1.00	4M	34	290
14	1.00		28	267
15	1.00		38	276
16	1.00	5M	35	296
17	1.00		23	284
18	1.00		54	290
19	1.00	6M	346	602
20	1.00		360	633
21	1.00		368	611
22	1.00	7M	385	645
23	1.00		386	665
24	1.00		430	678
25	1.00	8M	370	617
26	1.00		404	661
27	1.00		339	578
28	1.00	9M	392	657
29	1.00		374	617
30	1.00		418	657
31	1.00	10M	342	602
32	1.00		367	593
			312	546

MEA #1

11-30 a.m.

125/0dr + 50-200 µg/ml
MEA

Expt# : 1

Date/Time: 05/26/98

11-30 a.m.

12-00

TABLE-1

Expt. # : 1

Date/Time : 05/26/98; 11-30 a.m.

Tube #	Medium count for 10 ul (cpm)	Avg. cpm	dpm [cpm/0.7056]	μ Ci/ml (A _c) on counting [dpm/22200]	μ Ci/ml (A _o) on addition [A _c /e ^{-λt}]
1	-15, 7, -10	0	0	0	0
2	24, 0, 9	0	0	0	0
3	16, 8, -7	0	0	0	0
4	-6, -12, -2	0	0	0	0
5	-5, -17, 14	0	0	0	0
6	306, 320, 328	318	450.6	0.0203	0.0233
7	345, 346, 390	360.3	510.6	0.023	0.0264
8	330, 364, 299	331	469.1	0.0211	0.0242
9	352, 334, 378	354.6	502.6	0.0226	0.0260
10	302, 327, 272	300.3	425.6	0.0191	0.0220

05/14/98; 8:00 p.m.

0.8705

cpm for 300 ml Cells.

1	1.00		32	287
2	1.00		21	253
3	1.00		20	266
4	1.00	1c	35	258
5	1.00		31	282
6	1.00		34	246
7	1.00	2c	29	285
8	1.00		22	263
9	1.00		35	248
10	1.00	3c	26	239
11	1.00		22	261
12	1.00	4c	31	245
13	1.00		24	292
14	1.00		18	245
15	1.00		23	246
16	1.00	5c	23	283
17	1.00		28	268
18	1.00		556	815
19	1.00	6c	548	821
20	1.00		582	833
21	1.00		309	567
22	1.00	7c	318	541
23	1.00		265	512
24	1.00		377	649
25	1.00	8c	418	656
26	1.00		348	604
27	1.00		297	556
28	1.00	9c	307	571
29	1.00		346	575
30	1.00		668	917
31	1.00	10c	710	938
32	1.00		667	937
33	1.00		27	250
34	1.00		26	266

Expt Name : 125I Udr + 50-200 µg/ml
MEA

Expt # : 1

Date / Time : 05/26/98; 12:00
noon.

TABLE-2

Expt. # : 1

Date/Time : 05/26/98; 12-00

Tube #	Radioactivity for 300 ul cell suspension (cpm)	Avg. cpm	dpm [cpm/0.7056]	$\mu\text{Ci/ml (A)}$ on counting [dpm/666000]	$\mu\text{Ci/ml (A}_0)$ after 12 h incubation [$A/e^{-\lambda t}$]
1	-6, 9, 5	0	0	0	0
2	8, 3, -4	0	0	0	0
3	9, 0, -4	0	0	0	0
4	5, -2, -8	0	0	0	0
5	-3, -3, 2	0	0	0	0
6	530, 522, 556	536	759.6	0.00114	0.001296
7	283, 292, 239	271.3	384.5	0.00057	0.000656
8	351, 392, 322	355	503.1	0.00075	0.000859
9	271, 281, 320	290.6	411.9	0.00061	0.000703
10	642, 684, 641	655.66	929.2	0.00139	0.00158

05/15/98; 9-00 a.m

11 days + 3h

= 267

$$e^{-\lambda t}$$

$$= e^{-\frac{0.693 \times 267}{1440}}$$

$$= 0.8794$$

TABLE-3

Expt. # : |

Date/Time : 05/18/98

Tube #	Coulter count for 100 ul cell suspension	Avg. count	Cells/ml [Avg. count x 400]	pCi/cell uCi/ml x10 ⁶ Cells/ml]
1	480, 490, 499			0
2	392, 415, 442			0
3	311, 325, 309			0
4	375, 390, 392			0
5	350, 329, 341			0
6	425, 419, 433	425.6	170266.6	0.00761
7	280, 270, 262	270.6	108266.6	0.00605
8	355, 342, 352	349.6	139866.6	0.00614
9	285, 270, 272	275.6	110266.6	0.00637
10	475, 490, 411	458.6	183466.6	0.00861

TABLE-4

Expt #: 1

Date: 05/25/98

Colony Counts and Survival Fraction

Tube.dilution	Colony 1	Colony 2	Colony 3	Avg Colony	SF
1:2	85	82	87	84.66	
2:2	74	76	78	76	0.8977
3:2	69	75	72	72	0.8504
4:2	58	46	52	52	0.6142
5:2	48	52	40	46.6	0.5512
6:3	56	44	50	5.0	0.059
7:3	58	64	60	6.06	0.0797
8:3	194	160	184	17.9	0.249
9:3	120	140	116	12.5	0.241
10:3	65	62	63	6.33	0.1357

Conc. of
MEA
($\mu\text{g}/\text{ml}$)

DMF

50

0.90

100

1.67

150

1.67

200

1.56

Expt #1

DMF

10 NATIONAL
12-183
MADE IN U.S.A.

SF

0.1

0.01

Semi-Logarithmic
3 Cycles x 10 to the inch

0.002

0.004

0.006

0.008

0.010

0.012
psi/cell

0.014

